



# ExtravDose Modeling Theory

*for VARSKIN+ (V+) Version 2.0, software release: April 1, 2025*

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# Disclaimer

This presentation is an independent product of RCD and does not communicate the views or positions of the U.S. Nuclear Regulatory Commission (NRC).

# Training Progression for ExtravDose



Training on theory covers multiphysics mathematical models for fluid flow, activity concentrations, and dosimetry

← You are here.

Beginner training emphasizes Basic calculation

Intermediate training includes Advanced calculation features

ExtravDose discussions

# Outline for Training on Theory



## Part I

- Event Overview
- Variables to Consider
- Clinical Insights for Dose Modeling in Recent Literature

## Part II

- Multiphysics Modeling Overview
- Extravasated Fluid Flow in Tissue
- Activity Concentration in Tissue

## Part III

- Photon Dosimetry
- Electron Dosimetry
- Alpha Particle Dosimetry

## Part I

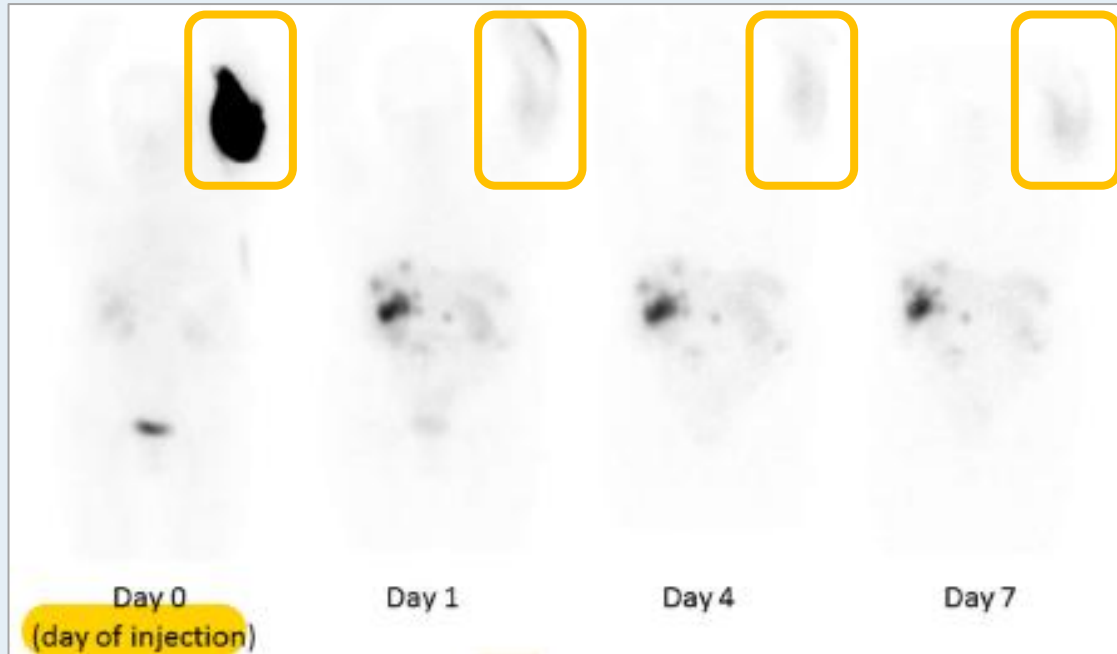


# – Extravasation dosimetry module



Extravasation occurs when radiopharmaceutical intended for the bloodstream leaks into surrounding tissue.

Temporary condition that elevates absorbed radiation dose to healthy tissue without a medical benefit



## OBJECTIVE

Develop models for a computational tool (software) to calculate 3D dose distributions in tissue with interfaces and flexibility appropriate for radionuclide extravasations

Arveschoug *et al.* *EJNMMI Research* (2020) 10:68  
<https://doi.org/10.1186/s13550-020-00658-6>



# What matters for modeling?



- Physical and chemical characteristics of the radiopharmaceutical
- Administration flow rate and duration (gravity method, infusion pump, injection)
- Biological composition and structure at the site of extravasation (tissue matrix)
- Physical changes to the site (elevation of site, warm compress)
- Biophysical movement and dissipation of extravasated fluid (flow & removal)
  - Volume of tissue affected by extravasated fluid
  - Extravasated fluid concentrations in tissue over time
- Radioactivity extravasated (initial source term)
- Radioactive decay, emission type and energy

# Multiphysics aspects addressed in 3 models

Flow Model

Movement of radioactive extravasated fluid in tissue

Activity Concentration  
Model

Activity concentration in tissue over time in three dimensions

Dosimetry Model

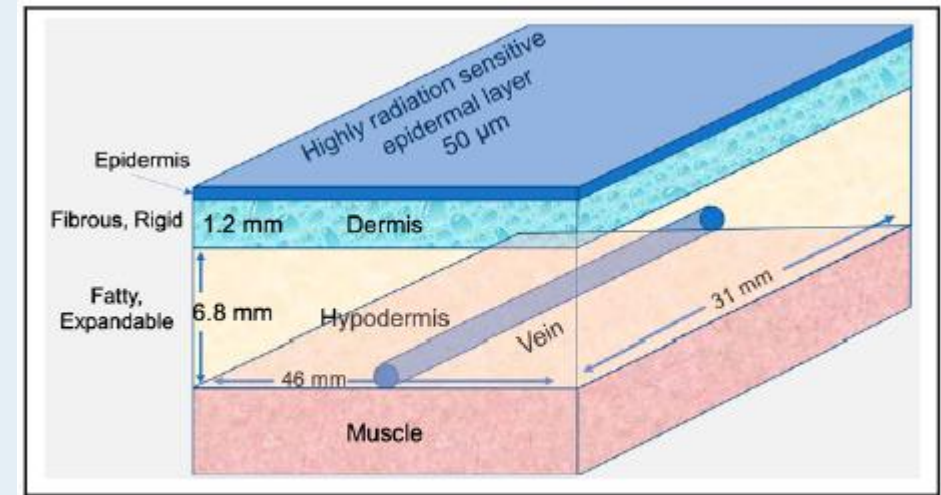
Local tissue dose over time in three dimensions



# Characterization of PET Extravasation (2 of 3)



“The dermis is a largely dense connective tissue, consisting primarily of collagen and elastin surrounded by glycosaminoglycans (highly polar water-binding molecules) that enable collagen fibers to retain water; this water is not freely exchangeable. For purposes of our dose calculations, hypodermis-to-dermis radioactivity concentration ratios of 10:1, 5:1, and 2:1 were simulated, but the results from the 10:1 simulations are likely closest to reality.”



**FIGURE 1.** Schematic of skin anatomy with primary dimensions used in Monte Carlo absorbed dose calculations.

Sutherland et al. (2023) Multicenter Evaluation of Frequency and Impact of Activity Infiltration in PET Imaging, Including Microscale Modeling of Skin-Absorbed Dose. *J Nucl Med* 64: 1095-1101. doi:10.2967/jnumed.123.265891.

# Characterization of PET Extravasation (3 of 3)



## Patient-specific micro-dosimetry analysis

- 1.2 mm dermis
- 6.8 mm subcutaneous layer (from CT images)
- Radioactivity remaining at injection site, 0.19% injected activity
- Volume of tissue infiltrated, 3.6 cm × 2.1 cm



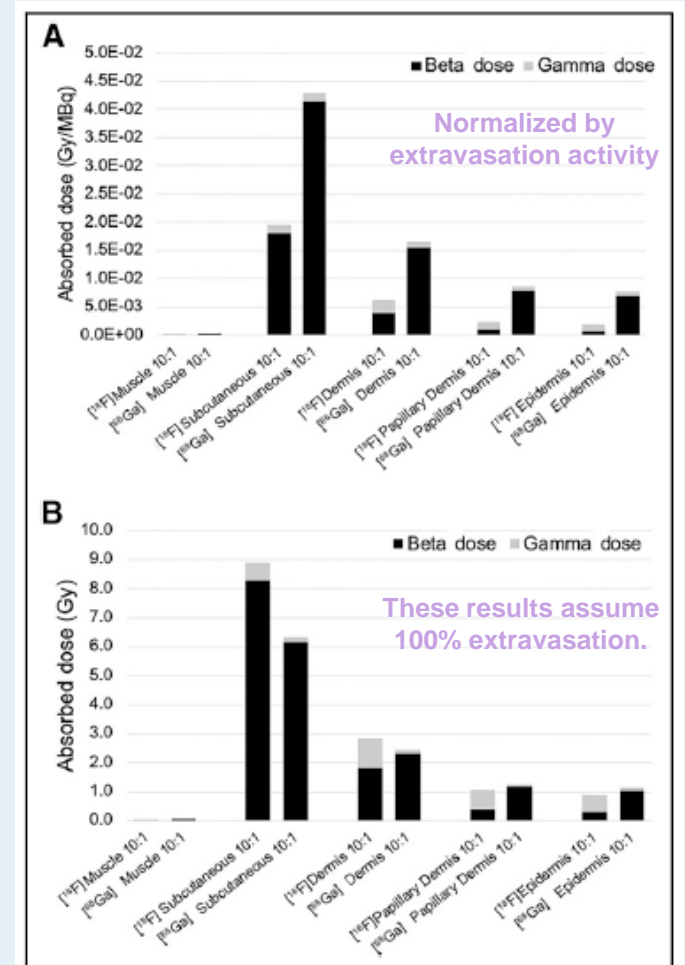
Patient E  
true infiltration  
forearm

## MCNP 6.2 modeling of absorbed dose to different tissue layers

- Assumed static activity distribution in hypodermis & dermis
- Assigned 10:1, 5:1, 2:1 hypodermis:dermis ratios for activity partitioning
- Authors preferred 10:1 based on animal experiments
- Effective half-lives: 25 min for  $^{68}\text{Ga}$  & 30 min for  $^{18}\text{F}$  from literature

## Highlighted need for quantitative measurements of activity partitioning between subcutaneous tissue & dermis

Sutherland et al. (2023) Multicenter Evaluation of Frequency and Impact of Activity Infiltration in PET Imaging, Including Microscale Modeling of Skin-Absorbed Dose. *J Nucl Med* 64: 1095-1101. doi:10.2967/jnumed.123.265891.



**FIGURE 5.** (A)  $\gamma$ - and  $\beta$ -contributions to absorbed dose for each sub-anatomy per megabecquerel infiltrated for  $^{18}\text{F}$  and  $^{68}\text{Ga}$ . (B) Relative  $\gamma$ - and  $\beta$ -contributions to absorbed dose for full infiltration of  $^{18}\text{F}$  and  $^{68}\text{Ga}$ .

# Biological clearance of $^{18}\text{F}$ -FDG from extravasation site

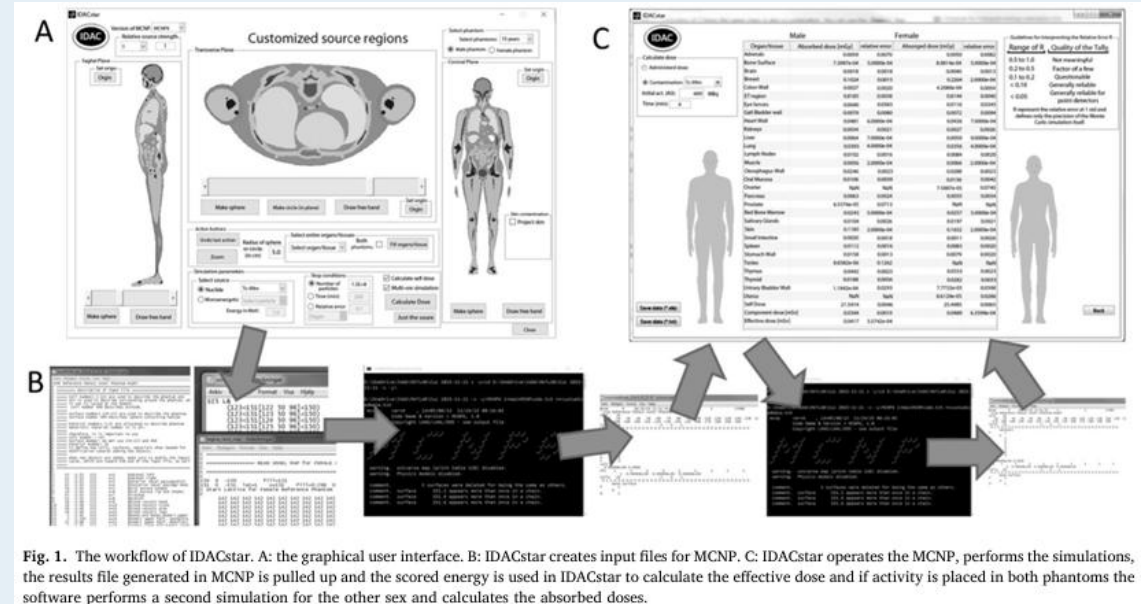


Ören et al. (2017) IDACSTAR: A MCNP application to perform realistic dose estimations from internal or external contamination of radiopharmaceuticals. *Radiation Protection Dosimetry*. 174(3): 406-411. doi:10.1093/rpd/ncw221.

For the extravasation, the effective dose was estimated to be 1.6 mSv assuming that all the activity being absorbed within the site of the extravasation. The activity in the extravasation area was estimated from the clinical images to be 90 MBq of  $^{18}\text{F}$ -FDG 87 minutes after the 370 MBq administration. Assuming a bi-exponential retention of the activity from the extravasation site to the blood, a biological half-time of 70 minutes was estimated. The effective dose for the more realistic case, where a fraction of 0.39 of the total number of disintegrations was in the extravasation volume, was estimated to be 4.5 mSv.

- Activity estimated from clinical images
- Bi-exponential retention at extravasation site
- $^{18}\text{F}$ -FDG biological half-time ~ 70 min

Andersson et al. (2022) IDACstar2.0: A MCNP application to perform realistic dose estimations from internal or external contamination with radiopharmaceuticals. *Radiation Physics and Chemistry*. <https://doi.org/10.1016/j.radphyschem.2021.109957>.



Graphical user interface creates input files for MCNP.

Papers address effective (whole-body) doses, not yet optimized for extravasation dose to local tissue.

# Extravasation modeling from PET/CT imaging (1 of 3)

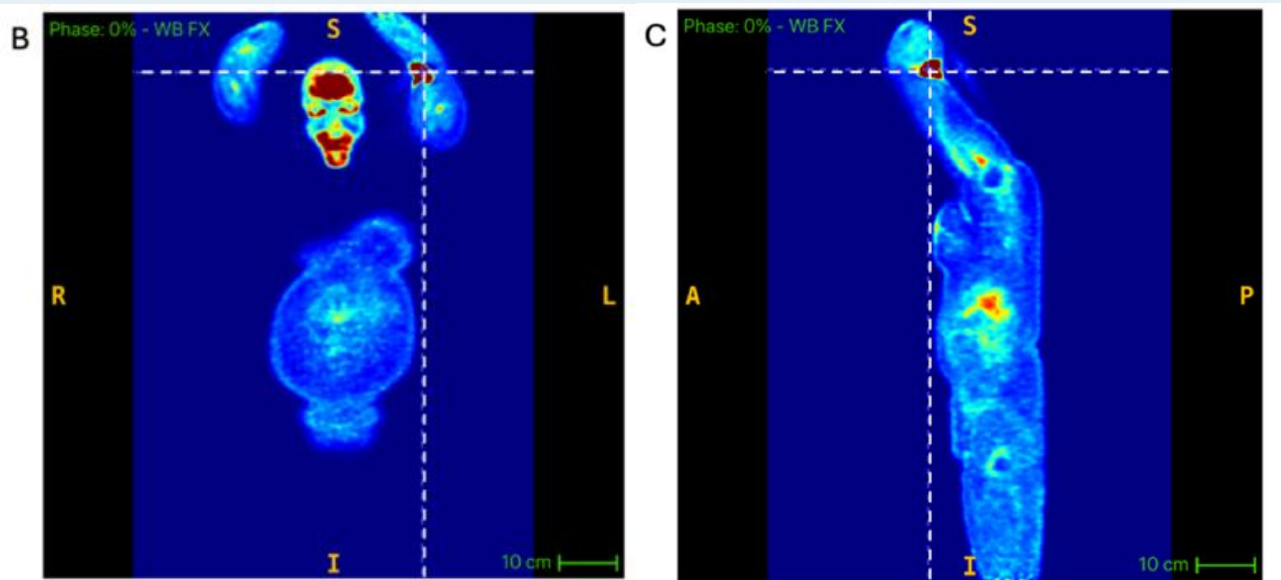


Fig. 1. ITK-SNAP [Paul et al., 2006] CT views of the injected patient. PET-CT images (A: axial, B: coronal, C: sagittal) showing the distribution of  $^{18}\text{F}$ -FDG radiopharmaceutical, with a notable accumulation in the extravasation region (e.g. dashed line points to the Region Of Interest, ROI).

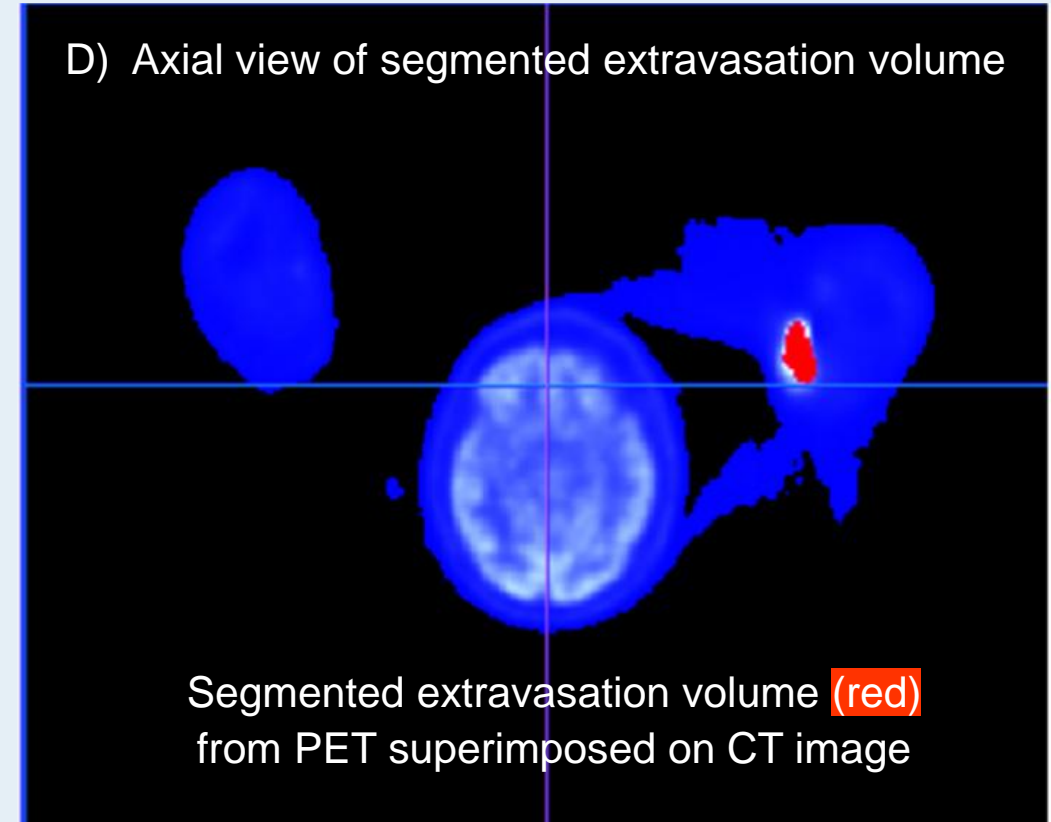


Fig. 2. Build-up phases for the generation of the FE model with Synopsis® Simpleware™ Medical. Panel A) Axial, B) Coronal, C) Sagittal CT views with evidence of the implemented segmentation of the tissues; ochre, skin; green, adipose tissue; red, muscle; pearl, bone. D) Axial view of the segmented extravasation volume superimposed on the CT image from the PET analyses, colors chosen to make evident the ROI. The legs were excluded from the FE model used for transport simulations obtaining a more manageable computational model, while preserving details of the of the ROI and critical organs in the upper body portion. (For interpretation of the references to color in this figure legend, the reader is referred to the Web version of this article.)

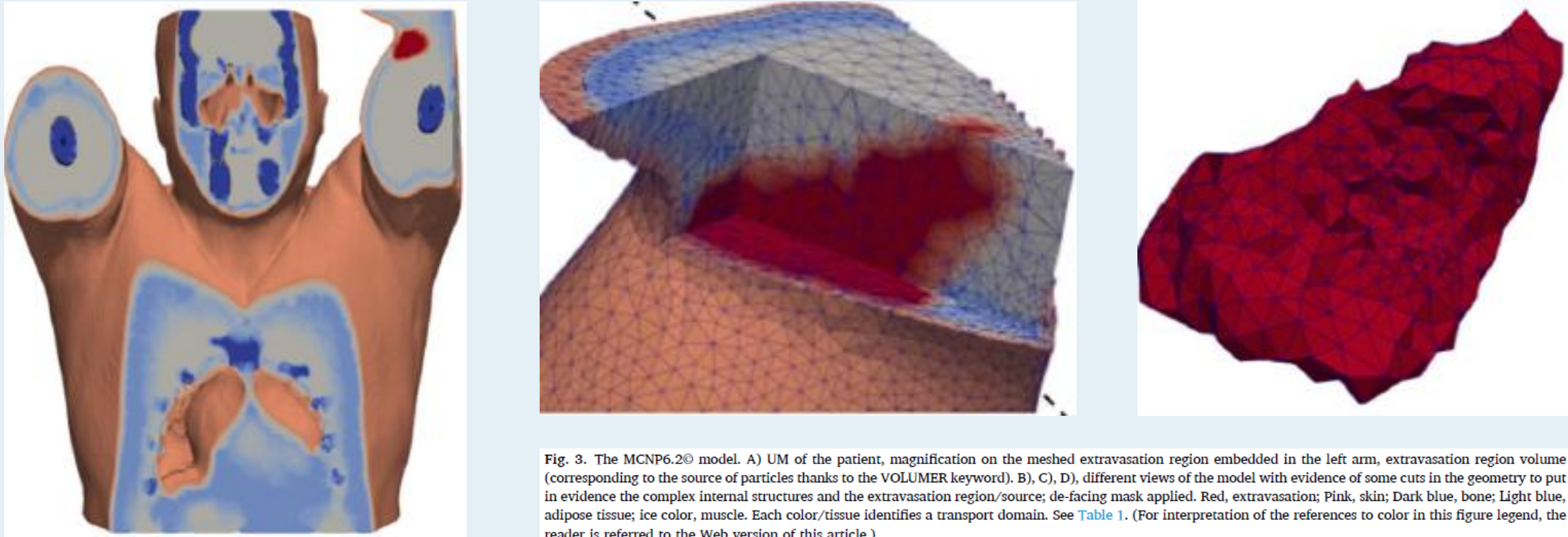


Fig. 3. The MCNP6.2© model. A) UM of the patient, magnification on the meshed extravasation region embedded in the left arm, extravasation region volume (corresponding to the source of particles thanks to the VOLUMER keyword). B), C), D), different views of the model with evidence of some cuts in the geometry to put in evidence the complex internal structures and the extravasation region/source; de-facing mask applied. Red, extravasation; Pink, skin; Dark blue, bone; Light blue, adipose tissue; ice color, muscle. Each color/tissue identifies a transport domain. See Table 1. (For interpretation of the references to color in this figure legend, the reader is referred to the Web version of this article.)

- Digital twin with four million spatial elements & related nodes for MCNP 6.2 dose modeling
- Incorporated  $^{18}\text{F}$ -FDG (fluorodeoxyglucose) effective half-life & mixing with blood
- Model imagery appears to show infiltration in dermis, hypodermis, and portion of adjacent muscle

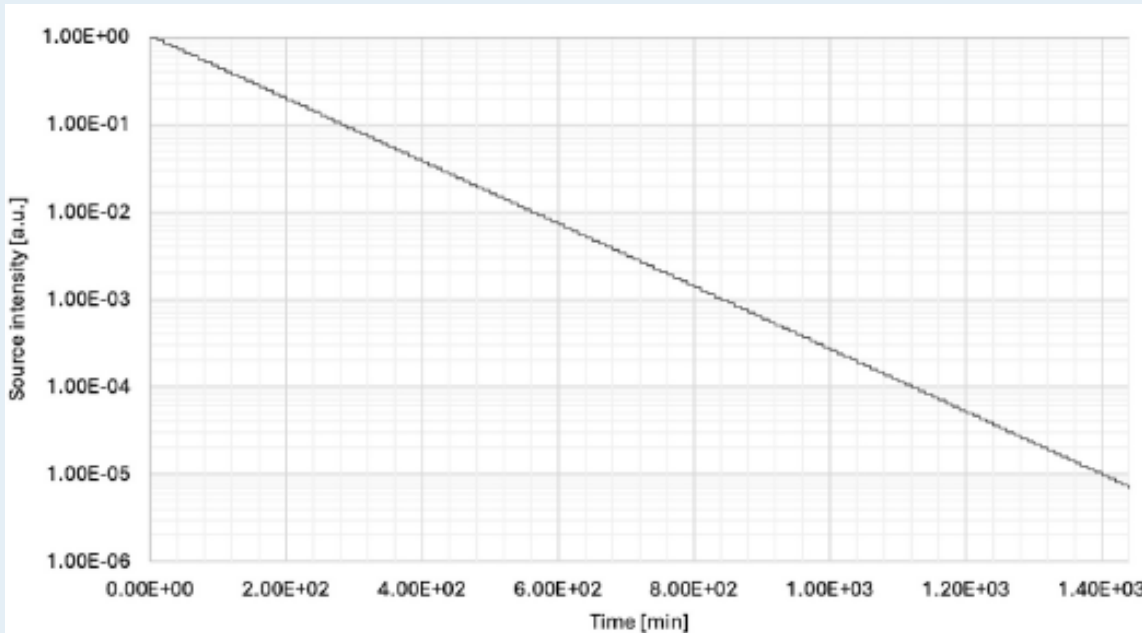
Isolan et al. (2025) Extravasation Modeling from a PET/CT Imaging for a Possible Digital Twin Approach. *Radiation Physics and Chemistry*. <https://doi.org/10.1016/j.radphyschem.2025.113080>.

# Extravasation modeling from PET/CT imaging (3 of 3)

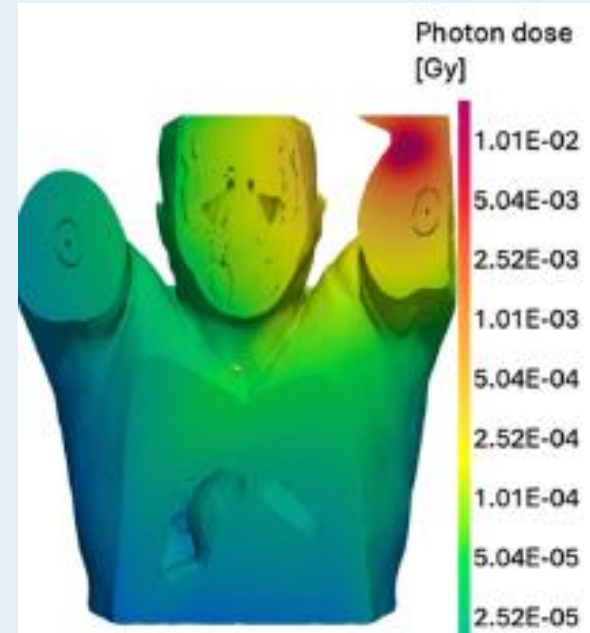


Applied effective half-life from literature to total radioactivity in infiltrated volume over time

Effective half-life ~ 84 min from slope (below)



Patient was injected with 227 MBq  $^{18}\text{F}$ -FDG  
228 mGy dose to extravasation volume



(from Figs. 4 & 5) Isolan et al. (2025) Extravasation Modeling from a PET/CT Imaging for a Possible Digital Twin Approach. *Radiation Physics and Chemistry*. <https://doi.org/10.1016/j.radphyschem.2025.113080>.

# $^{99m}\text{Tc}$ -HDP effective half-life at extravasation site

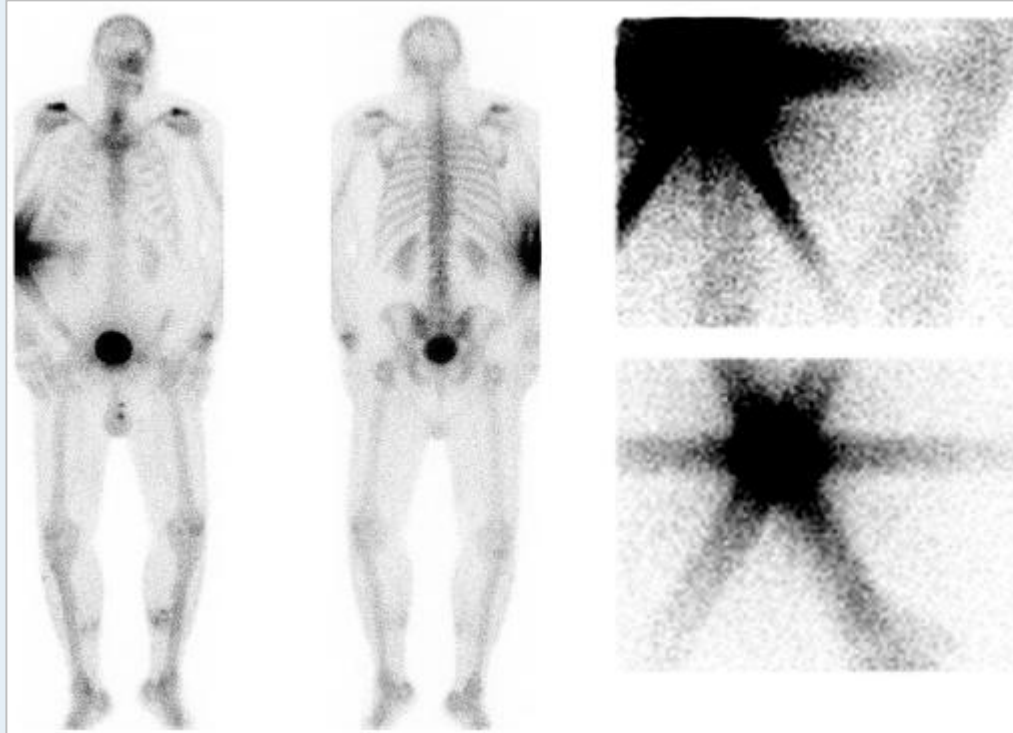


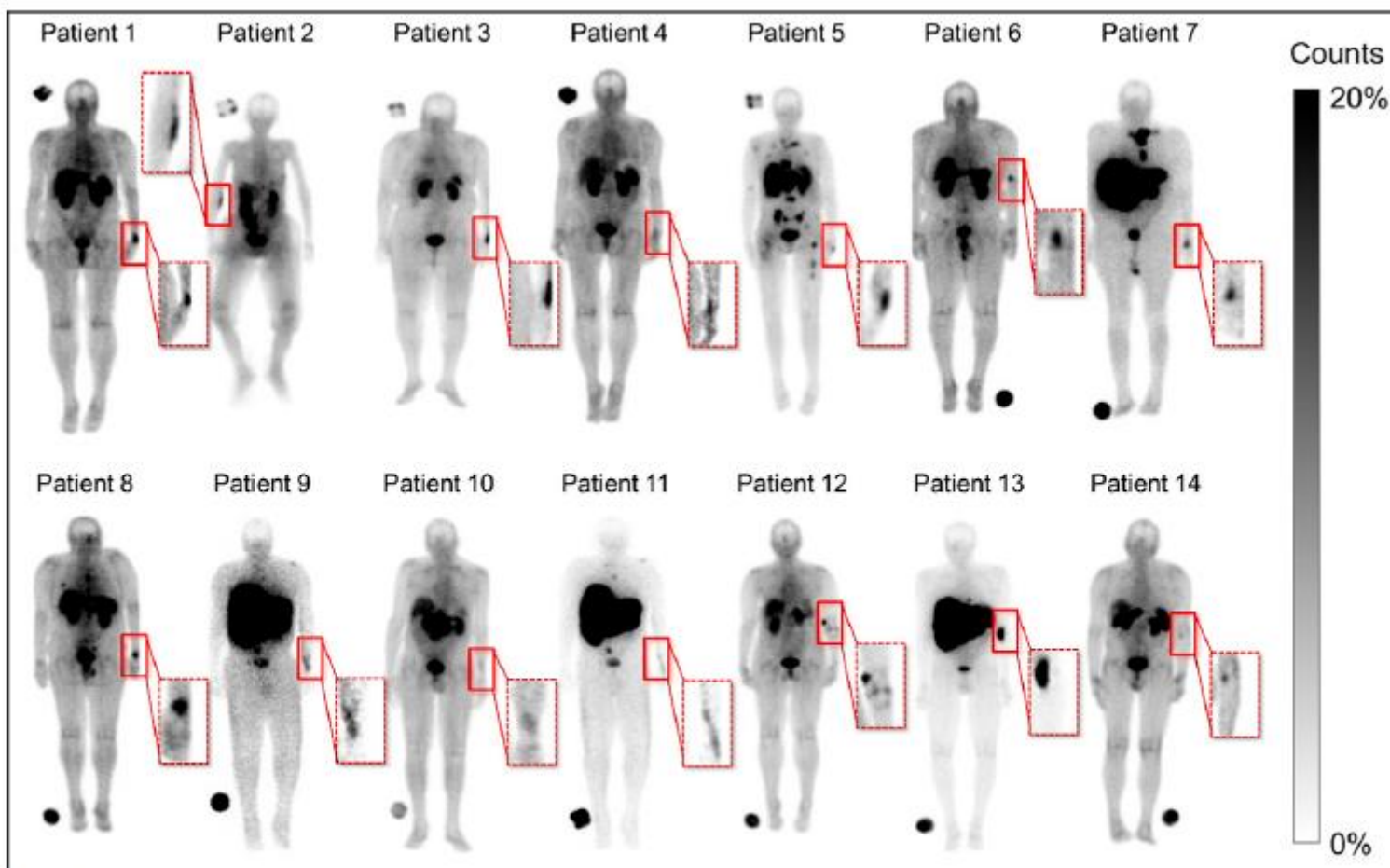
FIGURE 1 High activity at the site of extravasation in the right elbow is illustrated on anterior and posterior (AP) total-body planar scintigraphy images 4 h post-extravasation (left); and anterior images of the (AP) planar imaging of the right elbow injection site used for consequent dosimetry calculation, at 30 min (top right) and 4 h (bottom right).

- $^{99m}\text{Tc}$ -HDP effective half-life ~ 90 min
- Relatively simple modeling assumptions
- Local tissue doses not summarized here

A 50-year-old male patient presented with pain and dysfunction 1 year after a left-sided distal radius fracture. In order to better diagnose and treat the patient, 871 MBq [ $^{99m}\text{Tc}$ ]Tc-HDP (volume 6 mL, body weight 130 kg) was administered to perform three-phase bone scintigraphy.<sup>1</sup> After intravenous injection, dynamic imaging showed no visual activity at the wrist. This prompted the technician to suspect an extravasation. This extravasation was subsequently investigated further using planar imaging of the injection site (Figure 1).

Doornhof et al. (2024) Treatment of [ $^{99m}\text{Tc}$ ]Tc-hydroxy-diphosphonate ([ $^{99m}\text{Tc}$ ]Tc-HDP) extravasation using hyaluronidase. *Pharmacol Res Perspect.* 2024;12:e1232. <https://doi.org/10.1002/prp2.1232>.

# Extravasation in $^{177}\text{Lu}$ -DOTATATE therapy



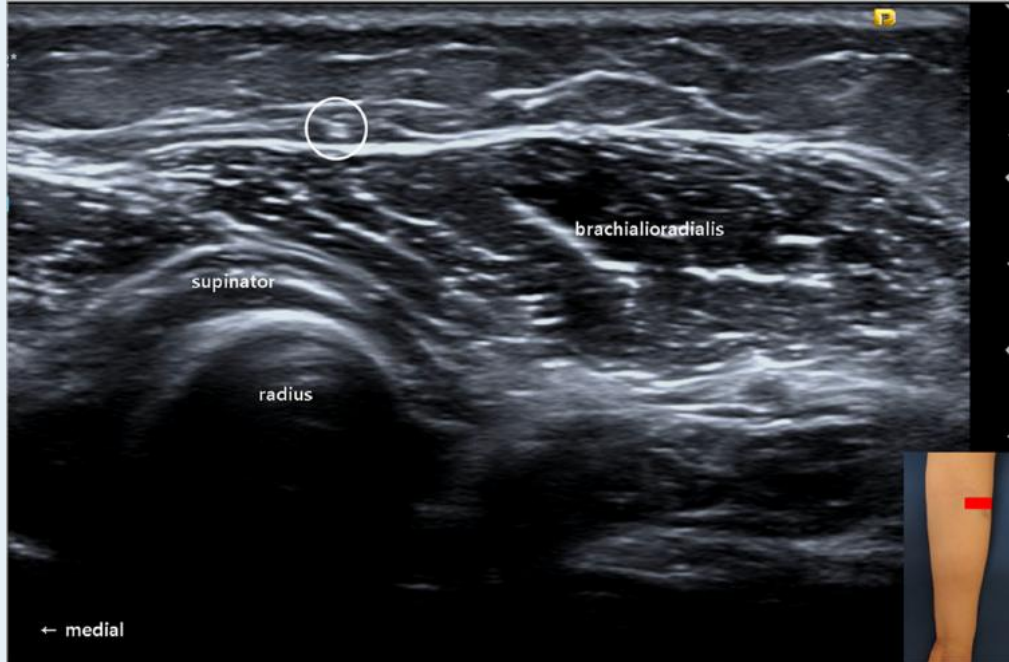
**FIGURE 1.** Planar gamma camera images (conjugate view geometric mean) of 14 patients with identified extravasations (highlighted in red dashed box) acquired using Philips XCT (patients 1–4) and Siemens Symbia 1 and 2 SPECT/CT systems (patients 5–14). All patients, except patient 2, had infusion in left arm. Gray scale was narrowed to 20% to enhance visualization of extravasations.

- Whole-body planar imaging performed 3-4 h after injection
- Extravasation occurs in hypodermis
- Homogeneous activity distribution assumed
- No patients retained more than 1% at injection site at time of scan
- No adverse skin reactions found within days to weeks
- Rapid dispersal from concurrent saline infusion & small molecule pharmacokinetics

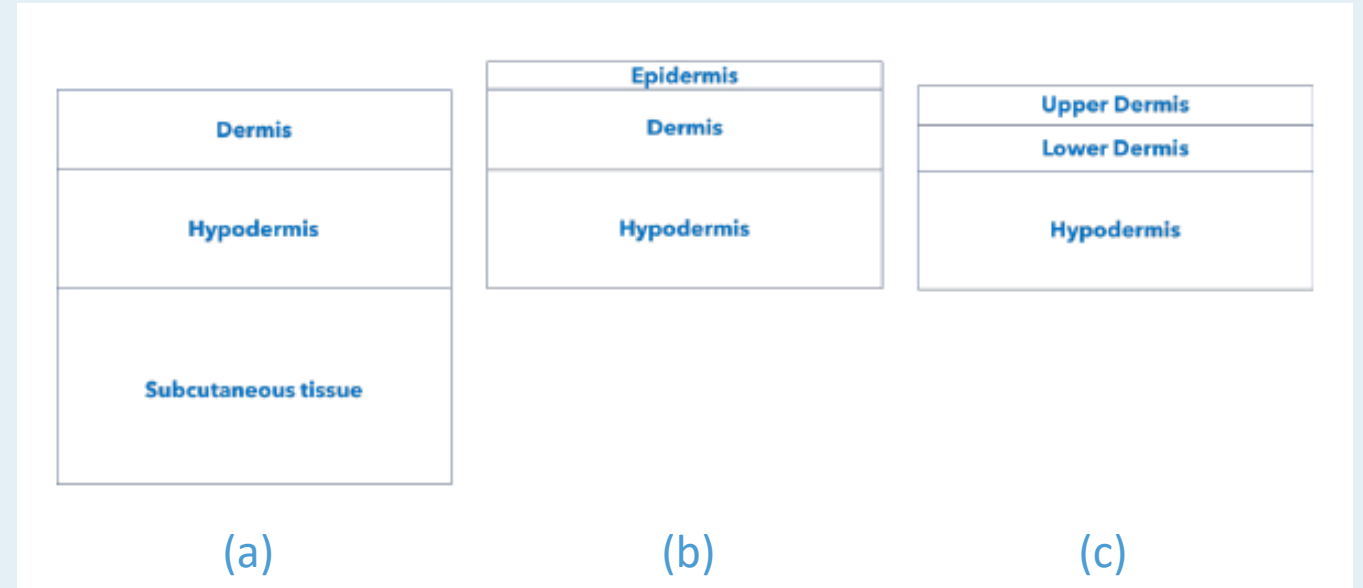
Kayal et al. (2025) Extravasation Frequency of [ $^{177}\text{Lu}$ ]Lu-DOTATATE: Insights and Implications Derived from 1,314 Cycles of Treated Patients—A Single-Site Analysis. *J Nucl Med* doi:10.2967/jnumed.124.269411. June 26, 2025.

## Part II

# Flexible tissue layering



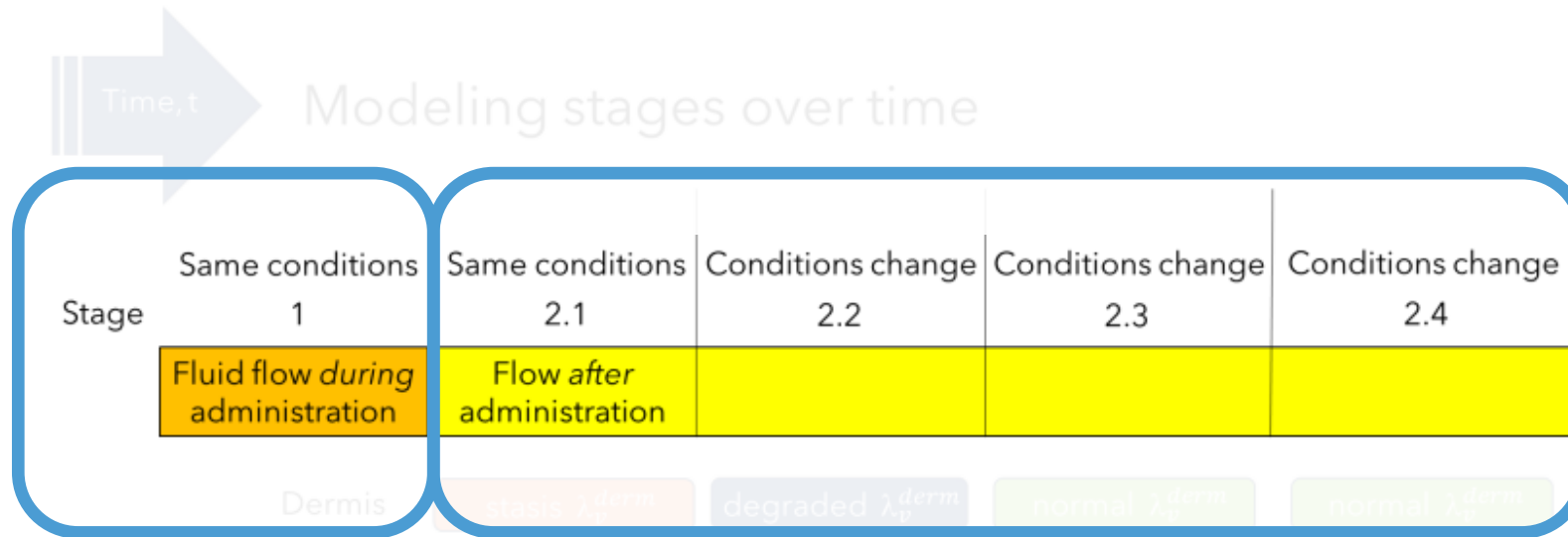
Kwak and Chang (2020) Compression of the lateral antebrachial cutaneous nerve due to leakage of iron after an intravenous iron infusion. *Diagnostics* 2020, 10, 516. Figure 1(C).



accommodates

- (a) Deep infiltration without appreciable extravasation in epidermis
- (b) Dermal extravasation with explicit infiltration into the epidermis
- (c) Confined extravasation within multiple dermal layers

# Flow primarily driven by pressure



## STAGE 1

If option is selected by user,  
External driving force to flow

This equation for confined flow

2D lateral calculation initially  
+ vertical fluid distribution

## STAGE 2

No external driving force

Darcy flow with hydrostatic pressure from fluid in tissue

3D calculation

# Discuss each model



Flow Model

Activity Concentration  
Model

Dosimetry Model

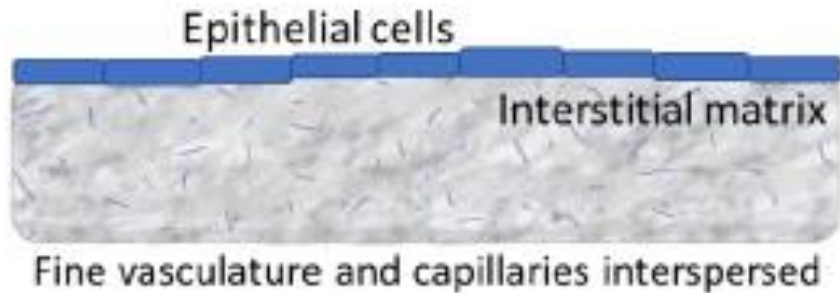
Starting with ...

Flow Model Overview

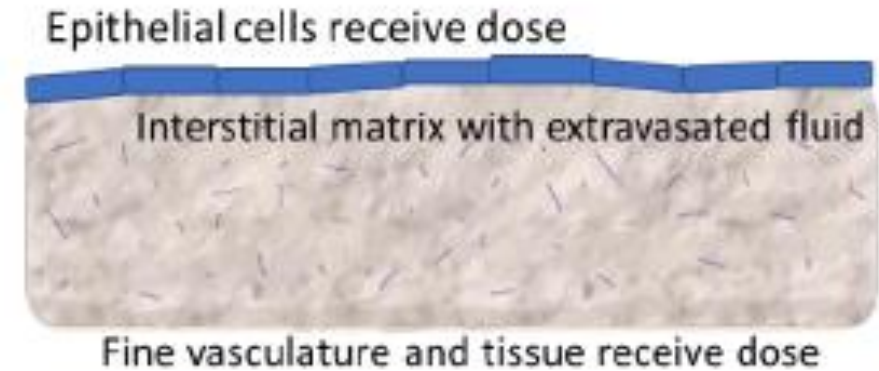
# Homogeneous tissue matrix

Interstitial components include fine vascular bathed in extracellular fluid.

before extravasation



after extravasation



In initial model, fluid movement is tracked in a stationary grid *without* tissue shape changes.

## Flow in Stage 1

External driving force to flow

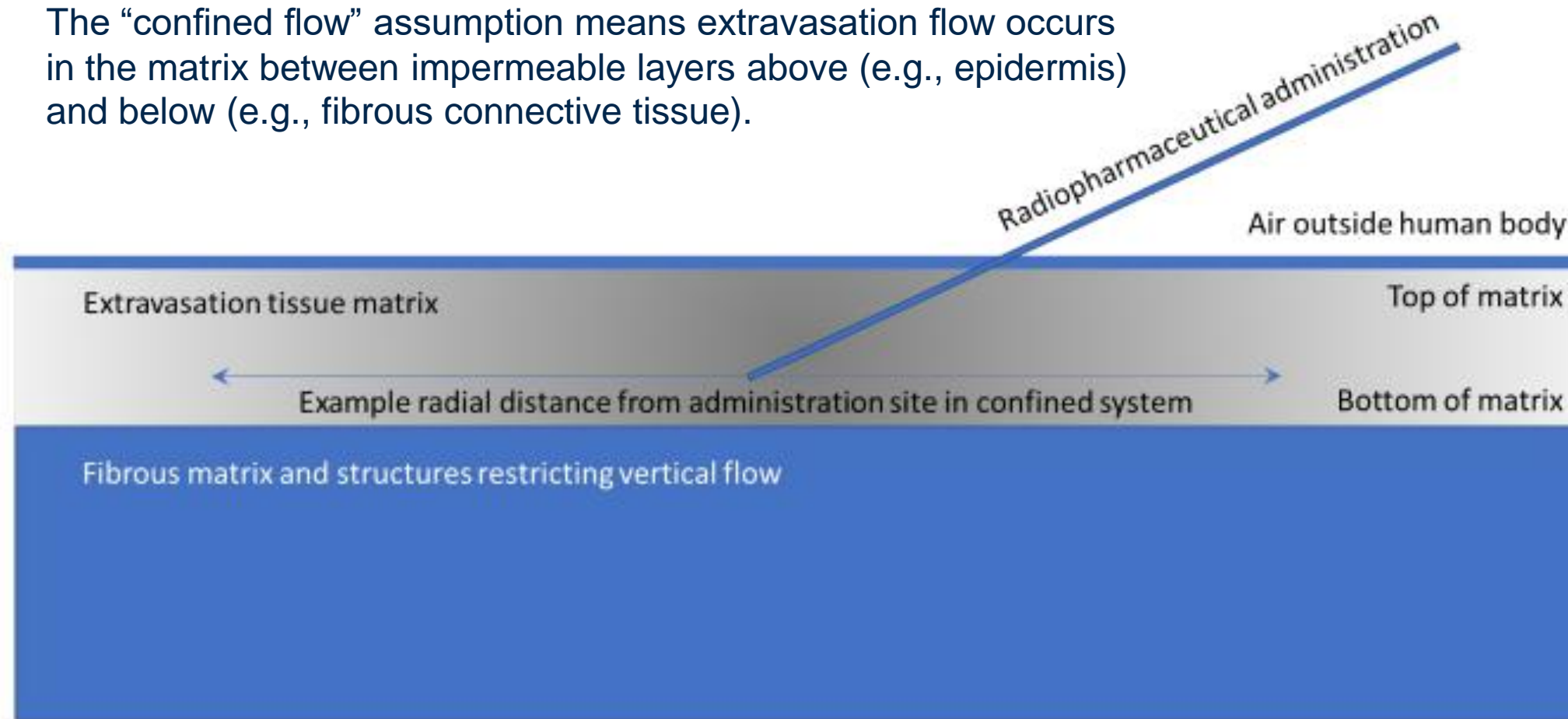
This equation for confined flow

2D lateral calculation initially + vertical fluid distribution

# Confined flow during administration in Stage 1



The “confined flow” assumption means extravasation flow occurs in the matrix between impermeable layers above (e.g., epidermis) and below (e.g., fibrous connective tissue).



# Theis Equation utilized for Stage-1 flow modeling



Hydrostatic pressure head [cm] as function of space and time

$$p(r, t) = \frac{Q}{4\pi T} E_1 \left( \frac{S}{4 T} \frac{r^2}{t} \right)$$

Exponential integral definitions

$$E_n(z) = z^{n-1} \int_z^{\infty} \frac{e^{-x}}{x^n} dx$$

$$E_1(z) = \int_z^{\infty} \frac{e^{-x}}{x} dx$$

- $Q$  Extravasation volumetric flow rate [ $\text{ml h}^{-1}$ ]
- $T$  Effective tissue transmissivity for extravasation lateral flow [ $\text{cm}^2 \text{h}^{-1}$ ]
- $S$  Storage coefficient for extravasated fluid in tissue [unitless]
- $r$  Radial distance from administration site [cm] and
- $t$  Elapsed time during radiopharmaceutical administration [h]

Storage coefficient is a relatively insensitive parameter for extravasation; therefore  $S = 0.02$  (unitless default value).

# Darcy Flow & Transmissivity



Darcy flow velocity [ $\text{cm s}^{-1}$ ] depends on pressure differentials and properties of the flow matrix

$$v = - \left( \frac{T}{h} \right) \left( \frac{\Delta p}{\Delta r} \right)$$

$h$       **Effective** thickness of tissue for extravasation flow [cm]

$\Delta p$       Hydraulic pressure gradient [cm] and

$\Delta r$       Distance over which the pressure gradient applies [cm]

$T = K h$       “Effective” accounts for spatial heterogeneity within the tissue matrix

$T$       **Effective** tissue transmissivity for extravasation lateral flow [ $\text{cm}^2 \text{h}^{-1}$ ]

$K$       **Effective** hydraulic conductivity for a homogeneous tissue matrix [ $\text{cm h}^{-1}$ ]

# Effective Transmissivity for Tissue Layers



$$T = \frac{\sum_{i=1}^m h_i T_i}{h}$$

Calculated as a weighted sum of transmissivities for individual layers  $i$

$T$	Effective transmissivity for lateral flow in the tissue matrix [ $\text{cm}^2 \text{h}^{-1}$ ]
$h_i$	Thickness of $i^{\text{th}}$ layer in the tissue matrix receiving extravasated fluid [cm]
$T_i$	Transmissivity for lateral flow in the $i^{\text{th}}$ layer of the tissue matrix [ $\text{cm}^2 \text{h}^{-1}$ ]
$i$	Index representing one of multiple layers [unitless] and
$m$	Total number of layers in the tissue matrix [unitless]

# Infiltrating Fluid Influences Local Transmissivity



To implement lateral flow, transmissivity is adjusted for combined effects of

- Hydraulic conductivity increases in fluid-filled tissue and
- Potentially greater flow heights

$$T = T_0 \cdot x^{\left(\frac{V_k + V_l}{2 s^3}\right)} \left(1 \frac{\text{cm}^3}{\text{ml}}\right)$$

Users can specify  $T_0$  and  $s$  in advanced calculations.

- $T_0$  Nominal transmissivity of the tissue layer without extravasation [ $\text{cm}^2 \text{h}^{-1}$ ]
- $x$  Base of the power relationship [unitless default value of 2 is assumed]
- $V_k$  Extravasated fluid volume in voxel  $k$  [ml]
- $V_l$  Extravasated fluid volume in voxel  $l$  [ml]
- $s$  Voxel side length [cm]

# Lateral Extravasated Fluid Flow during Administration



For a constant volumetric flow rate of the radiopharmaceutical during administration (Stage 1)

- Quasi-steady-state conditions for lateral extravasation flow would be reached within ~5 min
- Simplification is made to remove the time dependence from the Theis equation

Radial pressure gradient from the Theis equation yields a quasi-steady-state velocity profile

$$v = \frac{T}{h} \left[ \frac{p(r_0) - p(r_1)}{r_1 - r_0} \right]$$

Radial flow velocity solved at numerous radial positions



Bonta et al. (2011) Extravasation of a therapeutic dose of  $^{131}\text{I}$ -metaiodobenzylguanidine: Prevention, Dosimetry, and Mitigation. *J Nucl Med* 52: 1418-1422.

# Extravasated Fluid at Radial Locations Apportioned to Layers Based on Transmissivity



Extravasated fluid volume per unit tissue volume at various radial positions in each layer,  $\delta_{r_j,i}$  [ml cm<sup>-3</sup>]

Calculated by dividing the infiltrating extravasated fluid volume,  $V_{r_j}$ , by the original tissue volume of that layer at radial position  $r_j$

Volume of extravasated fluid [ml] in radial ring  $j$  is apportioned to layer  $i$

$$\delta_{r_j,i} = \left( \frac{V_{r_j}}{h_i A_{r_j}} \right) \left( \frac{T_i}{\sum_{i=1}^m T_i} \right)$$

index  $j \in \{0, 1, \dots, n\}$  for radial rings

index  $i \in \{1, \dots, m\}$  for tissue layers

Tissue volume [cm<sup>3</sup>] for layer  $i$  within radial ring  $j$

$A_{r_j}$  Cross-sectional area for the  $j^{\text{th}}$  radial ring [cm<sup>2</sup>]

## Flow in Stage 2

No external driving force

Darcy flow with hydrostatic pressure from fluid in tissue

3D calculation

## Stage 2 Addresses Flow after Administration Ceases



Parameters of state for the computational cell

$$P_k \quad P'_k \quad E_k \quad V_k \quad V'_k$$

Generic cell index =  $k$

Symbol	Units	Parameter Description
$P_k$	cm	Hydrostatic pressure head at start of time step
$P'_k$	cm	Hydrostatic pressure head at end of time step
$V_k$	ml	Volume of extravasated fluid at start of time step
$V'_k$	ml	Volume of extravasated fluid at end of time step
$E_k$	cm	Elevation head relative to administration site

Residual hydrostatic pressure from extravasated fluid temporarily trapped in tissue

$$P_k = 5.70 \left[ 1 - e^{-2.92 \left( \frac{V_k}{s^3} \right)} \right]$$

$V_k$  is the extravasated fluid volume in the computational cell at position  $k$  [ml]  
 $s^3$  represents the volume of one 3D computational grid for Stage 2 flow modeling [cm<sup>3</sup>]

## Stage-2 Lateral Flow in 3D grid



$$\Delta V_{k \leftarrow l} = \frac{\Delta t}{60} \left\{ T[(P_l + E_l) - (P_k + E_k)] \left( 1 \frac{\text{ml}}{\text{cm}^3} \right) + D \left( \frac{V_l - V_k}{s^2} \right) \right\}$$

- $\Delta V_{k \leftarrow l}$  Extravasated fluid volume flowing laterally into cell  $k$  from adjacent cell  $l$  during the time step [ml]  
 $\Delta t$  Time step [min]  
 $s$  Lateral distance between adjacent cells, side length for uniform cubic grid [cm]  
 $P_k$  Hydrostatic pressure head in cell  $k$  [cm]  
 $P_l$  Hydrostatic pressure head in cell  $l$  [cm]  
 $E_k$  Elevation head for cell  $k$  relative to the site of administration [cm]  
 $E_l$  Elevation head for cell  $l$  relative to the site of administration [cm]  
 $V_k$  Extravasated fluid volume in computational cell  $k$  [ml]  
 $V_l$  Extravasated fluid volume in computational cell  $l$  [ml]  
 $T$  Transmissivity for lateral extravasation flow within a tissue layer [ $\text{cm}^2 \text{h}^{-1}$ ]  
 $D$  Diffusivity of extravasated fluid in tissue [ $\text{cm}^2 \text{h}^{-1}$ ]

## Stage-2 Vertical Flow in 3D Grid



$$\Delta V_{k \leftarrow l} = \frac{\Delta t}{60} \left[ U \left( \frac{s}{d} \right) (P_l - P_k) \left( 1 \frac{\text{ml}}{\text{cm}^3} \right) + D \left( \frac{V_l - V_k}{d s} \right) \right]$$

$\Delta V_{k \leftarrow l}$  Extravasated fluid volume flowing vertically into cell  $k$  from adjacent cell  $l$  during the time step [ml]

$U$  Transmissivity of vertical flow across tissue layers [ $\text{cm}^2 \text{h}^{-1}$ ]

$d$  Effective vertical distance between adjacent cells for flow modeling [cm]

$$d = s + \left( \frac{V_k + V_l}{2s^2} \right) \left( 1 \frac{\text{cm}^3}{\text{ml}} \right)$$

Large volumes of infiltrating fluid increase the effective distance over which hydrostatic pressure difference acts for vertically adjacent computational cells (voxels)

$V_k$  Extravasated fluid volume for cell  $k$  [ml]

$V_l$  Extravasated fluid volume for cell  $l$  [ml]

$s$  Voxel side length [cm]

# Vascular-Lymphatic Removal of Extravasated Fluid



Vascular-lymphatic removal of extravasated fluid from the  $k^{\text{th}}$  computational cell (voxel) during each time step

$$V_k^- = \frac{\lambda_v \Delta t}{60} V_k$$

$\lambda_v$  Relative vascular-lymphatic removal rate [ $\text{h}^{-1}$ ]

In the current implementation, a single relative vascular-lymphatic removal rate of  $0.15 \text{ h}^{-1}$  is used in Stage 2.

If a warm compress event is included as a timeline input, a removal rate of  $0.25 \text{ h}^{-1}$  is used for that time period.

# Limb Elevation Influences Relative Elevation Head Calculation

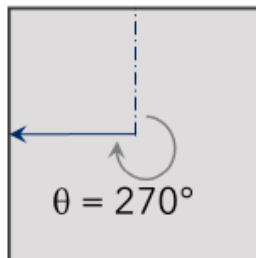


$$E_k = r_k \cos(\theta - \theta_k) \sin(\varphi)$$

- $E_k$  Relative elevation head for the  $k^{\text{th}}$  computational cell [cm]
- $r_k$  Radial distance of the  $k^{\text{th}}$  computational cell [cm]
- $\theta_k$  Polar angle for the computation cell arranged within the grid [degrees]
- $\theta$  Angle for the lift direction [degrees]
- $\varphi$  Tissue lift angle [degrees]

User-specified angles for (left) polar angle  $\theta$  for lift direction and (right) azimuthal angle  $\varphi$  for lift magnitude

Polar angle determines direction



Azimuthal angle determines lift

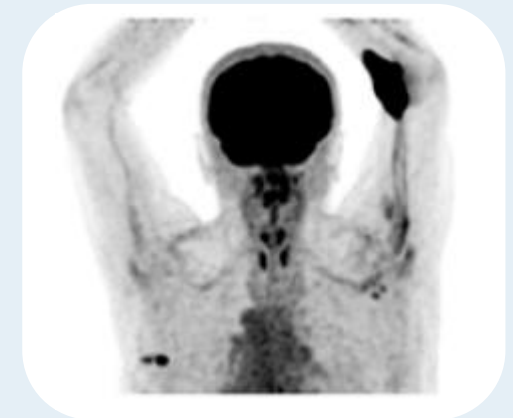
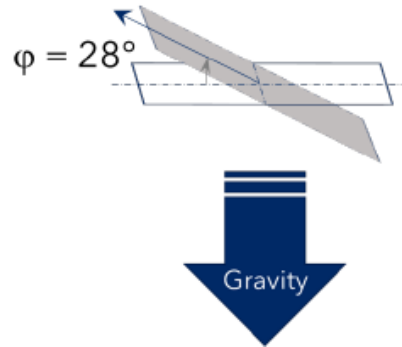


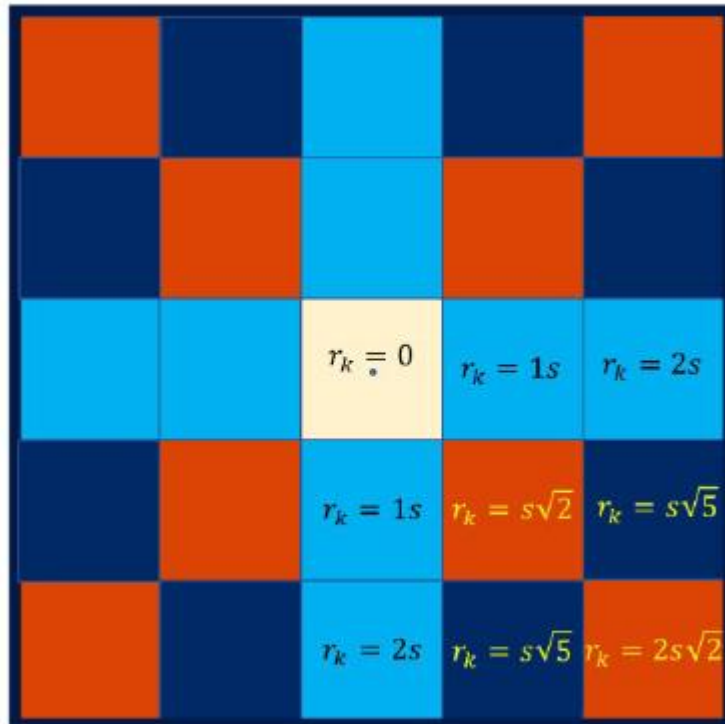
Fig. 3(C) in Iori et al. (2022)  
<https://doi.org/10.21203/rs.3.rs-2009242/v1>



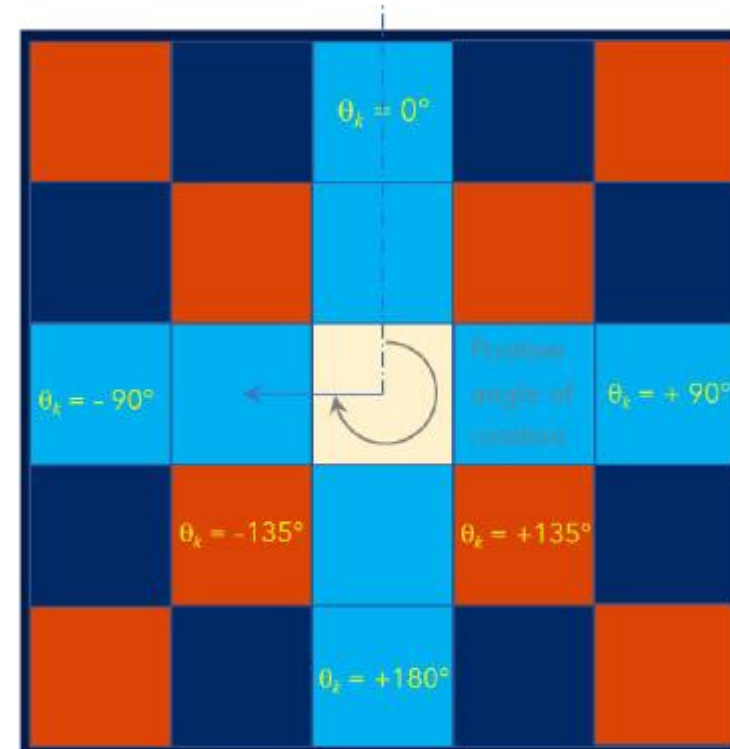
# Radial Distances for Elevation Head



$r_k$  is the cell center distance to center of grid



$\theta_k$  is the polar angle for lift direction



Radial distances and polar angles for a square grid with a side length  $s$

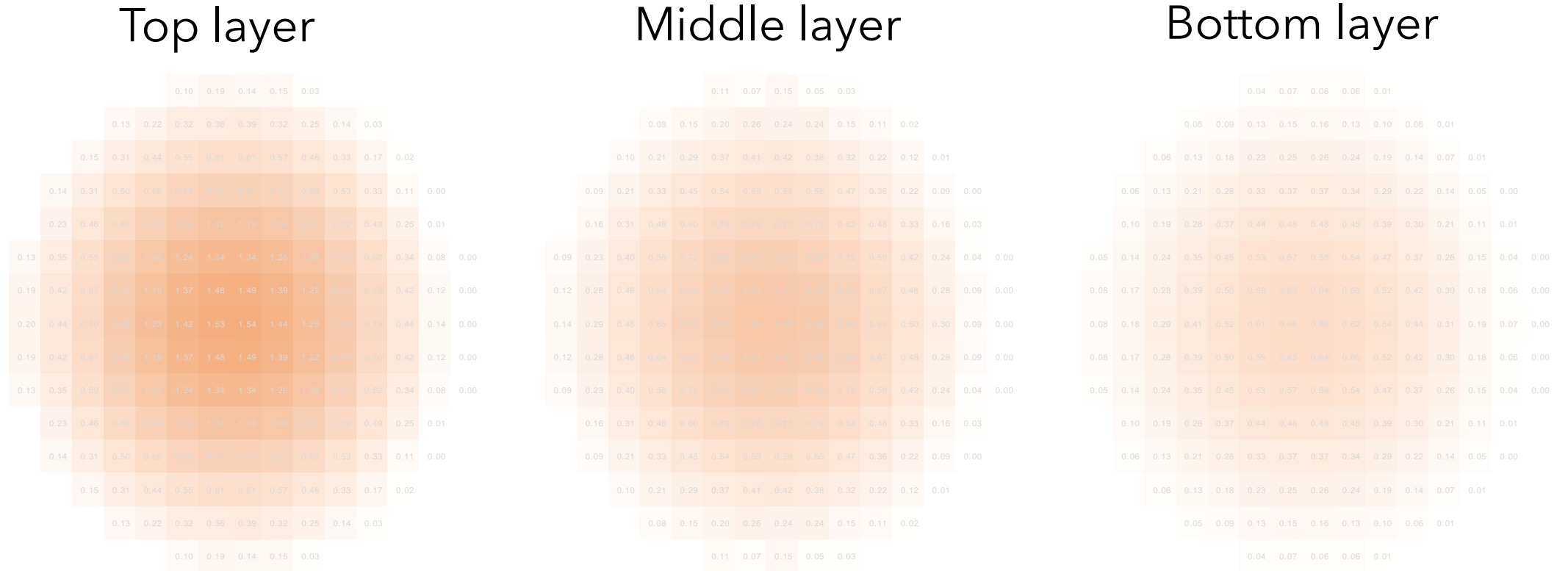
## Activity Concentration Model

# Activity Concentration Model



Extravasated  
fluid volume  
conc. [ml cm<sup>-3</sup>]

2.4  
2.2  
2.0  
1.7  
1.5  
1.2  
1.0  
0.7  
0.5  
0.2  
0.0



- Converts extravasated fluid in tissue (test flow result above) into activity per unit volume
- Applies 3D computational grid with unit tissue density that does not flex or deform
- Transfers spatial- & time-dependent **activity concentrations in tissue** to dosimetry model

# Activity Concentration Parameters & Constraint



- Utilizes radiopharmaceutical activity concentration in extravasated fluid
  - Unitless fraction for nonradioactive fluid in tissue displaced during extravasation, set to 0.33 (extracellular fluid fraction)
  - No additional inputs for this model
- Biological processes do not concentrate extravasated radioactivity in tissue
- Constraint limits activity concentration in tissue

$$C_{k,i}^* \leq C$$

$C_{k,i}^*$  Activity concentration in tissue [Bq cm<sup>-3</sup>]

$C$  Activity of radiopharmaceutical per unit extravasated fluid volume [Bq ml<sup>-1</sup>]

with 1 cm<sup>3</sup> = 1 ml

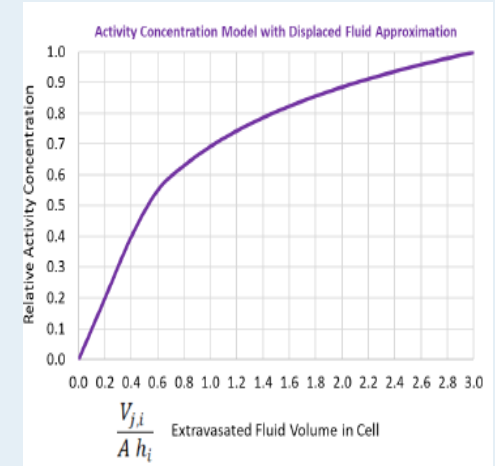
- Model considers dilution with nonradioactive fluid

# Activity Concentration in Tissue

based on extravasated fluid volume in computational cells (voxels)



$$C_{k,i}^*(t) = \begin{cases} F_{k,i}^* C e^{-\lambda_r t} & \text{if } F_{k,i}^* \leq F \\ \frac{1}{(1 + F_{k,i}^* - F)} F_{k,i}^* C e^{-\lambda_r t} & \text{if } F_{k,i}^* > F \end{cases}$$



$C_{k,i}^*(t)$  Activity concentration in tissue [Bq cm<sup>-3</sup>] at time  $t$  [min]

$C$  Activity of radiopharmaceutical per unit extravasated fluid volume [Bq ml<sup>-1</sup>]

$F_{k,i}^*$  Extravasated fluid at the location expressed as a fraction of the computational cell volume [unitless],

$$F_{k,i}^* = \left( \frac{V_{k,i}}{s^3} \right) \left[ \frac{ml}{cm^3} \right] \text{ where } 1 \text{ cm}^3 = 1 \text{ ml}$$

$V_{k,i}$  Extravasated fluid volume in a computational cell at the  $k^{\text{th}}$  location in the  $i^{\text{th}}$  layer, output from the extravasation flow model [ml]

$s$  Lateral distance between adjacent cells, side length for uniform cubic grid [cm]

$F$  Fraction of nonradioactive fluid in tissue that is displaced during extravasation,  $0 \leq F \leq 1$  [unitless]

$\lambda_r$  Radioactive decay constant, relative rate of removal [min<sup>-1</sup>]

## Part III: Dosimetry Model

## Extravasation Dosimetry in the Literature

- Generally, methods involve self-absorption assumptions in spheres of various size (i.e., “affected volume”)
- Modeling may include time-dependent removal of extravasated radioactivity from the affected volume
- The pharmaceutical (chemical structure) is very important for determining biological loss rates
- Deterministic threshold doses on the order of 2-20 Gy for erythema, epilation, edema, desquamation, blistering, ulceration, burns, and necrosis.

# Activity Model for Dose Estimation

## – Dosimetry assumptions

- biological outcome is deterministic
- homogeneous tissue ( $\rho = 1.1 \text{ g/cm}^3$ ) containing both source and target
- 3D grid structure of fine cubic cells with specified side length,  $s$
- gross volume of “affected tissue”  $W \times L \times D$
- concentration per cell as a function of time passed from transport model

## – Activity in a given voxel is proportional to radiopharmaceutical concentration

$$A_{k,i}(t) = C_{k,i}^*(t) \cdot s^3$$

$k^{\text{th}}$  location in the  $i^{\text{th}}$  layer

# Number of Dosimetry Calculations

- Considering contributions from photons, electrons, and alpha
- Generally, using point-kernel methods with two considerations:
  - energy self-absorption in each voxel
  - emissions from a source voxel to energy absorption in a distant target voxel
- A very large number of dose calculations are carried out:
  - for example, an affected volume of 20 x 40 x 1 calculational cells results in about 5.8M dosimetry calculations for each time step
  - In Phase 1
    - one time step for each simulated second
  - In Phase 2
    - the number of time steps is dependent on fluid velocity (Courant-number limited)
    - there could be tens-to-hundreds of time-steps per simulated minute

# Extravasation Photon Dosimetry



- The point-kernel method is implemented for photons
- For self-absorption and distant energy deposition
- Photon absorbed dose rate ( $\text{Gy s}^{-1}$ ) at distance  $r$ , is calculated as shown below

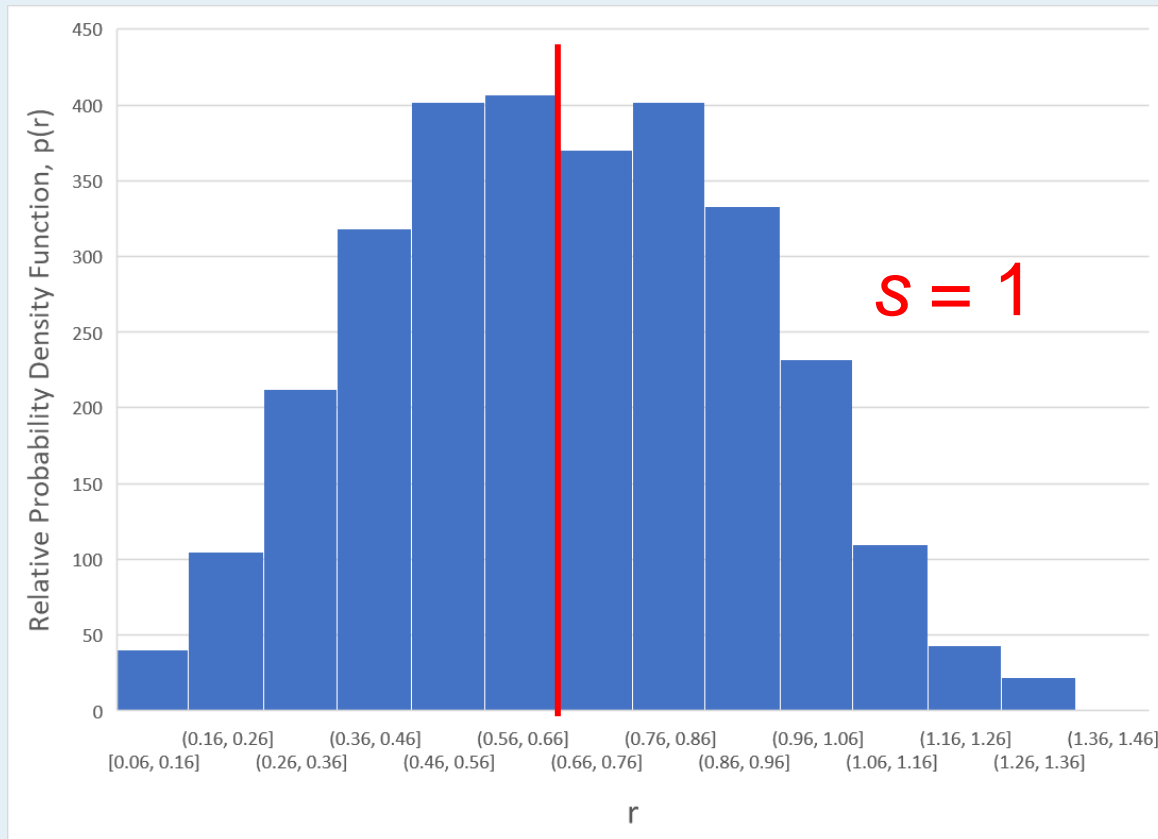
$$\dot{D}_\gamma(r) = \frac{k A Y E_0}{4\pi r^2} \left( \frac{\mu_{en}}{\rho} \right) e^{-\mu r}$$

$k$	unit conversion [ $1.6 \times 10^{-10} \text{ J g MeV}^{-1} \text{ kg}^{-1}$ ]
$A$	activity in the calculational cell [ $\text{nt s}^{-1}$ ]
$Y$	emission yield [ $\gamma \text{ nt}^{-1}$ ]
$E_0$	uncollided photon energy [ $\text{MeV } \gamma^{-1}$ ]
$r$	point-kernel distance [cm]
$\frac{\mu_{en}}{\rho}$	mass energy absorption coefficient [ $\text{cm}^2 \text{ g}^{-1}$ ]
$\mu$	linear attenuation coefficient [ $\text{cm}^{-1}$ ]

# Self-Absorption in the Source Voxel



Given a unit cube, we can randomly sample multiple combinations of source and target points. The distance between the two points is related to  $r$  (i.e., the probability of  $r$ ,  $p(r)$ ) in the point-kernel equation. The distribution of these random point kernel distances is referred to as a point-pair distribution.



Based on the histogram, the point-kernel distance used for cells that are both source and target is assumed to be two-thirds of voxel side length specified by the user (i.e.,  $r = \frac{2}{3}s$ ).

# Distant Voxels

Source (S) emissions represented at voxel center

Target (T) receptor represented at voxel center

Dose calculated by point kernel method using distance,  $r$

When source and target are different cells, distance between computational cells,  $r$ , with centroid 3D coordinates of  $(x_1, y_1, z_1)$  and  $(x_2, y_2, z_2)$ , is calculated as

$$r = \sqrt{(x_2 - x_1)^2 + (y_2 - y_1)^2 + (z_2 - z_1)^2}$$

With non-adjacent voxels, this point-kernel method works well, i.e., to within 10% of Monte Carlo volume-to-volume methods.

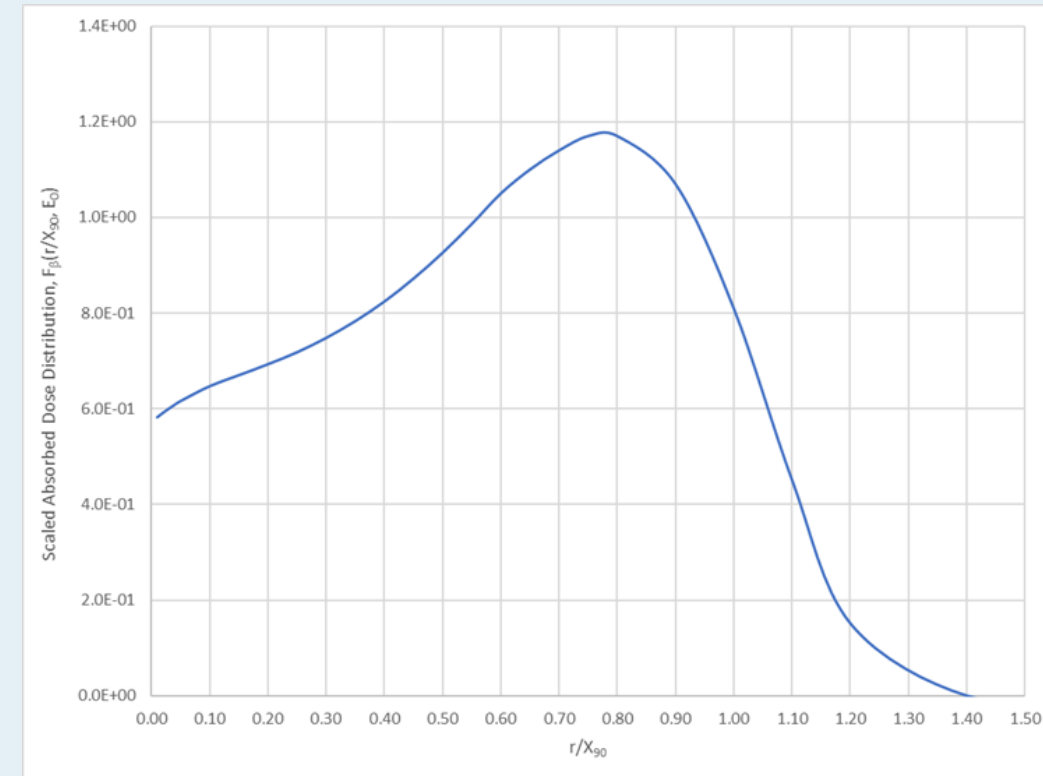
# Extravasation Electron Dosimetry

- Electron dosimetry methods utilize the SkinDose (V+) SADCALC routine with this point kernel:

$$\dot{D}_e(r) = k \frac{A Y \bar{E}_e F_e(\delta, E_o)}{\rho 4\pi r^2 X_{90}}$$

- With these additional parameters:

- $X_{90}$  = distance from source in which 90% of electron energy is deposited [cm]
- $F_e(\delta, E_o)$  = scaled absorbed dose distribution (SADD) [unitless]
- $\delta$  is the scaling index for point-kernel distance (i.e.,  $r/X_{90}$ )



## Similar to photons

- For instances where source and target are the same voxel, again the assumption applies:  $r = \frac{2}{3}s$
- In cases where the  $X_{90}$  is very short relative to the cubic side ( $X_{90} < \frac{s}{4.5}$ ), the electron energy is shown to be completely absorbed in the source voxel, thus:

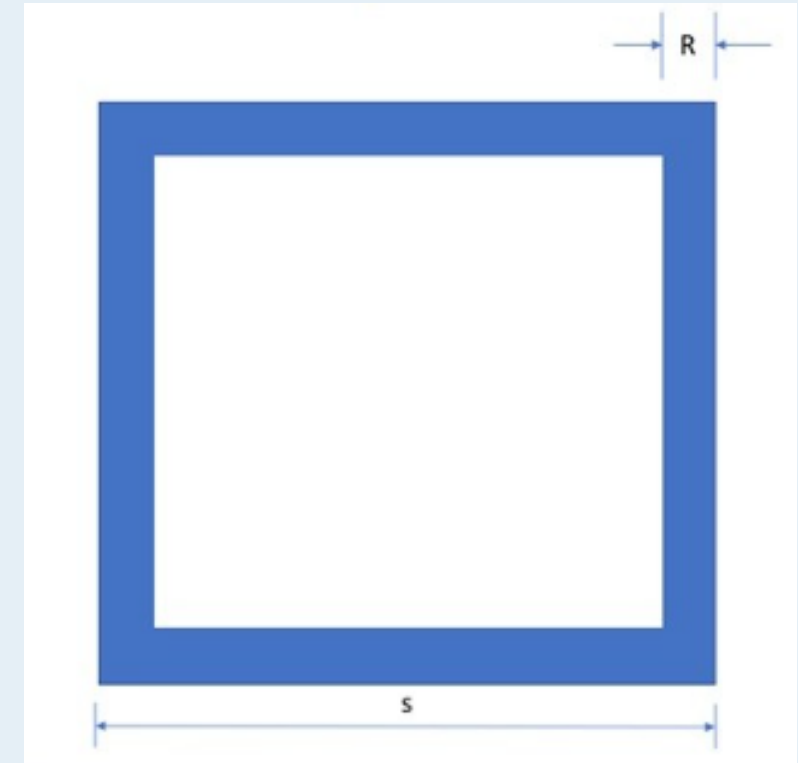
$$\dot{D}_e = k \frac{A Y \bar{E}_e}{\rho s^3}$$

# Extravasation Alpha Dosimetry

- Alpha particles travel no more than about 100 microns in tissue
  - recoil atoms also carry kinetic energy, but in even shorter distances
- Therefore, the point-kernel method is not appropriate
- An energy-balance method is used instead
- The majority of emitted alpha energy is absorbed in the source voxel (self absorption)
  - with a small fraction assumed to be absorbed in one of the six adjacent voxels
- Alpha dosimetry is dependent on alpha range
  - i.e., what fraction of energy from the source voxel enters an adjacent voxel

# Alpha energy leaving the source voxel

- Alpha emissions from a given source are uniformly distributed in a source voxel. Given the alpha range,  $R$ , and the voxel side dimension,  $s$ , the volume of the shell in the source voxel from which these alphas can escape to adjacent voxels is determined.
- For alpha emissions originating in that shell, we would expect roughly  $\frac{1}{2}$  to be direct into the voxel and the other  $\frac{1}{2}$  directed out of the voxel.
- The energy fraction transported to one of six adjacent voxels is calculated.
- This fraction is dependent on the source voxel dimension,  $s$ , and the alpha range,  $R$



# Alpha Dosimetry

– Alpha absorbed dose rate to the source voxel is:

$$\dot{D}_{\alpha S} = k \frac{A Y E_{\alpha}}{\rho s^3} \left(1 - \frac{3R}{s}\right)$$

– Alpha dose rate to each of the adjacent voxels is:

$$\dot{D}_{\alpha A} = k \frac{A Y E_{\alpha}}{\rho s^3} \left(\frac{R}{2s}\right)$$

For  $R = 0.005$  cm  
and  $s = 1$  cm,

$$\left(1 - \frac{3R}{s}\right) = 0.985$$

and

$$\left(\frac{R}{2s}\right) = 0.0025$$

## Main Component Highlights



Benjegerdes et al. (2017) Focal cutaneous squamous cell carcinoma following Radium-223 extravasation. Baylor University Medical Center Proceedings 30:1, 78-79, DOI: 10.1080/08998280.2017.11929538.

### Local Tissue Dose from Extravasation

#### – Flow Model

- Stage 1 during Radiopharmaceutical Administration
- Stage 2 after Radiopharmaceutical Administration

#### – Activity Concentration Model

#### – Dosimetry Model

### Presently not included in **ExtravDose** calculations

#### – Basal layer dose

#### – Decay progeny dose contributions

## Questions